



INTERNATIONAL JOURNAL
OF PHARMA AND BIO SCIENCES

Internationally Indexed Journal

Indexed in Chemical Abstract Services(USA),Index Copernicus ,Ulrichs Directory of Periodicals,Google scholar ,Cabi ,DOAJ ,PSOAR, EBSCO ,SCOPUS, EMBASE etc.



Rapid Publishing

The International Journal of Pharma and Bio Sciences (IJPBS) is an international journal published quarterly. The Aim of IJPBS is to publish peer reviewed research and review articles in less time without much delay in the developing field of Pharmaceutical and Biological sciences. One week from the date of manuscript submission author gets the decision of acceptance and if accepted the manuscript will be processed within 3 weeks (approx.) for publication.

Pharmaceutical Sciences

Branches

- Pharmaceutics
- Novel drug delivery sys
- Nanotechnology
- Pharmacology
- Pharmacognosy
- Analytical chemistry
- Pharmacy practice
- Pharmacogenomics
- Polymer sciences
- Biomaterial sciences
- Medicinal chemistry
- Natural chemistry
- Biotechnology
- Pharmacoinformatics
- Biopharmaceutics

Biological Sciences

Branches

- Biochemistry
- Biotechnology
- Bioinformatics
- Cell biology
- Microbiology
- Allied Sciences
- Molecular biology
- Neurobiology
- Cytology
- Pathology
- Immunobiology



Impact Factor 0.47*

**Refer Instruction to authors available at www.ijpbs.net*



Indexed In Elsevier Bibliographic Database
(SCOPUS, EMBASE and Sciverse)

Journal Home page
www.ijpbs.net

FOR INSTRUCTION TO AUTHORS VISIT
www.ijpbs.net

- FOR ANY QUERIES EMAIL TO
- editorofijpbs@yahoo.com
 - editorijpbs@rediffmail.com
 - prasmol@rediffmail.com



**PHYTOPHARMACOLOGICAL EVALUATION OF *SOLANUM SURATTENSE* BURN:
USED IN FOLK MEDICINES OF CHOLISTAN DESERT PAKISTAN.**

SOFIA ERAM^A, MAHMOOD AHMAD^B AND SHAFIA ARSHAD*^C

^{a,c}University College of Conventional Medicine, Faculty of Pharmacy and
Alternative Medicine, the Islamia University of Bahawalpur, Pakistan.

^bDepartment of Pharmacy, Dean Faculty of Pharmacy and Alternative Medicine,
the Islamia University of Bahawalpur, Pakistan.

ABSTRACT

Ethanollic extracts of aerial parts of *Solanum surattense* burn. (SS), were subjected to *in vivo* study, by means of serum biochemical parameters and histopathological observations, in order to verify its traditional use in Hepatobiliary disorders. All groups of rabbits received a single dose of CCl₄ subcutaneously on 7th day except group I which was served as normal control. Group II served as CCl₄ control. While groups III, IV and V were served as silymarin control, test group 1 and test group 2 respectively. Results showed that CCl₄ control group had raised levels of SGOT, SGPT and ALP significantly but TB level was not raised as compared to a normal control group. Pre-treatment with SS extract showed hepatoprotection as obvious by significant reinstatement of levels of SGOT, SGPT and ALP, while TB level was not changed significantly when compared with CCl₄ control group. SS extract was more effective as compared to both silymarin and SS extracts because SS extract only significantly reduced SGOT level but unable to restore SGPT, ALP and TB levels. Histopathological examination of the liver tissue was further corroborated these results. Thus the outcome of the present study supports the conventional believes on hepatoprotective effects of *Solanum surattense*.

KEY WORDS: *Solanum surattense*, Hepatoprotection, Carbon tetrachloride, Serum biochemical parameters, Histopathology of liver.



SHAFIA ARSHAD

University College of Conventional Medicine, Faculty of Pharmacy and
Alternative Medicine, the Islamia University of Bahawalpur, Pakistan.

INTRODUCTION

Cholistan desert is present on the Eastern side of the Punjab province, Pakistan (Baig et al., 1980). It surrounds thirty kilometers from Bahawalpur, Punjab, Pakistan and cover an area of 16,000 km² (Chouhan et al., 2002). The majority of plants grows in the desert has therapeutic properties and native people utilize these plants to treat various diseases (Shafi et al., 2001). *Solanum surattense* burm. (Family: Solanaceae) is one of those plants and is commonly known as "kanderi/kanderi wal". It is a herbaceous plant. The whole plant is used for medicinal purposes. In folk medicine, Decoction of its root is used to treat phlegmatic cough and fever. Whole plant decoction is very effective in skin diseases. Decoction of its fruit is given in bronchial asthma. Leaves juice with few seed of black pepper is a very useful remedy in joint pain. Whole plant decoction is given in case of jaundice (Qureshi, 2004). In some places, whole plant and fruit are used for food purposes (Nohara et al., 2007). Native people of Cholistan desert use this plant in hepatobiliary disorders. Experimental studies shows that SS possess antifungal activity against *A. fumigates*, *A. flavus*, *A. niger* and *C. albicans*. It also shows significant anti-microbial activity especially against *S. typhi*, *E. coli*, *P. aeruginosa*, *S. aureus* (Dabur et al., 2007). SS shows significant anti-inflammatory and anti asthmatic activities because of its steroidal constituents (Maiti et al., 1979). It is also known to possess Strong anti-proliferative activity against many cancer cells, anti-herpes and hypoglycemic activities (Nohara et al., 2007). However, to the best of our knowledge, no previous work has been published on hepatoprotective effects of this plant. Therefore, the present study was aimed to evaluate the hepatoprotective activity of aerial parts of SS against CCl₄-induced hepatotoxicity.

MATERIALS AND METHODS

Ethanol, CCl₄, Formalin, Diagnostic kits (SGPT, SGOT, ALP, and TB), Xylene, Paraffin wax,

Eosin, Hematoxylin and Canada balsam. The subsequent chemicals were purchased from Merck, Darmstadt, Germany. Silymarin and Pentothal sodium was obtained from Abbott Laboratories, Pakistan. Olive oil was from P. Sasso, Italy. All chemicals of analytical grade were used. *Solanum surattense* (aerial parts) was collected from Cholistan Desert and authenticated by a Taxonomist. Plant material was dried under shade, cut into small pieces and then subjected to grinding. The coarse powder (3000 g) of plant material was macerated in 9 L of ethanol for approximately 15-20 days with frequent shaking. The extract was filtered and marc left behind. The extract was concentrated under reduced pressure on Rotary evaporator until a semisolid residue is obtained. Marc was further extracted under the same conditions twice. These semisolid residues collected from extraction were combined and evaporated to dryness by vacuum at a temperature below 60 °C. At the end a dark brownish green solid residue was obtained and approximate yield was 310.2gms. For convenient administration, the dry extract powder was encapsulated after weighing. Healthy rabbits of either sex (local breed), weighing from 1.5-2 kg was purchased from local markets. They were kept in the animal house of Faculty of Pharmacy and Alternative Medicine, the Islamia University of Bahawalpur. The animals were maintained at standard housing conditions and fed standard pellet diet and water ad libitum. All procedures were performed according to the institutional animal Ethics Committee's approval. Hepatotoxicity was induced subcutaneously by CCl₄ at a dose of 0.75 ml/kg body weight, suspended in olive oil (1:1). The animals were randomly divided into five groups, containing ten rabbits in each. CCl₄ was injected 30 minutes after drug administration, on the 7th day of the 8 days study period to all the groups except group I which was served as normal control and received only normal saline. Group II-V received the following treatments from 1st to 7th day of the study.

Group II CCl₄ control (normal saline at 5 ml/kg/day)

Group III Silymarin control (100 mg/kg/day)

Twenty-four hours after administration of CCl₄, blood samples (3ml) from all the five groups were drawn from Jugular vein by sterile disposable syringe. Blood samples were allowed to coagulate at room temperature for 45 min into sterile dry centrifuge tubes. Serum was separated by centrifugation at 2500 rpm for 15 min and subjected to biochemical analysis. Merck diagnostic kits and UV-VIS Spectrophotometer (U2020 IRMECO, Germany) were used to measure serum glutamate oxaloacetate transaminase (SGOT), serum glutamate pyruvate transaminase (SGPT), serum alkaline phosphatase (ALP) and total Bilirubin (TB). 7 rabbits per each group were selected randomly for histopathological examination. Histopathological assessment was done according to the standard method (Humason, 1979). The pathological changes of fatty liver and degeneration of liver cells were graded as given below;

- Grade 0 (Normal): Normal liver morphology; hepatocytes with round nucleus centrally with homogenous cytoplasm, flat endothelial cells around central vein and sinusoid.
- Grade +1 (Mild degree): 1-2 hepatocyte rows around central vein showed; hepatic cell

Group IV *Solanum surattense* extract (500 mg/kg/day)

Group V *Solanum surattense* extract (750 mg/kg/day)

degeneration along with necrosis (loss of nucleus), less injury of endothelial cells around central vein, less fat vacuoles in hepatocytes.

- Grade +2 (Moderate degree): Some hepatocyte rows around central vein showed; swelling, intracytoplasmic vacuolar degeneration in centrilobular, midzonal and periportal areas endothelial cells around the central vein more damage than level +1 more fat vacuoles in hepatocytes than level +1.
- Grade +3 (Severe degree): 3-4 hepatocyte rows around the central vein demonstrated; hepatocytic degeneration and necrosis, degeneration cells including centrilobular, midzonal and periportal areas (diffuse intracytoplasmic vacuolar degeneration), endothelial lining of central vein showed more cell damage, increased fat vacuoles in hepatocytes than level +2, marked focal necrosis. The results were presented as Mean \pm Standard error of the means (S.E.M). Multiple comparisons were performed by student's *t*-test. Differences were considered statistically significant when $P < 0.05$.

RESULTS AND DISCUSSION

Administration of CCl₄ (0.75 ml/kg, s.c.) produced a significant increase in serum enzyme levels, namely SGOT, SGPT and ALP. However, TB level was remained unchanged when compared with normal control. The protective action of SS aerial parts extracts on CCl₄ induced hepatotoxicity are summarized in Table 1. Pretreatment with SS extract (500 mg/kg), caused a significant reduction in the levels of SGOT, SGPT and ALP while TB level remained unchanged, as compared to CCl₄ control group. Pretreatment with SS extract (750 mg/kg), produced less significant results

than SS extract (500 mg/kg) and decrease only SGOT level with no restorative effect on SGPT and ALP levels as compared to CCl₄ control. SS extract (500 mg/kg) was also significantly more effective than Silymarin. Histopathological changes after 24 h of CCl₄-induced liver injury included hepatocytes necrosis, inflammatory cell infiltration, fatty degeneration, hydropic degeneration, vacuole generation and microvascular steatosis. Administration of the SS extract (500 mg/kg) significantly preserved the almost normal hepatocellular architecture from the damaging effects of CCl₄ as compared to both Silymarin (100 mg/kg) and SS extract (750 mg/kg). The scoring of histological damage is

presented in Table 1. Induced acute hepatocellular damage is frequently used indicator to date for the assessment

CCl₄-induced acute hepatocellular damage is frequently used indicator to date for the assessment of the hepatoprotective potential of drugs or medicinal flora and their extracts, both via *in vivo* and *in vitro* techniques (Weber et al., 2003). The hepatic damage is evident by increase in the level of released cytoplasmic transaminases (SGOT and SGPT), alkaline phosphatases (ALP), in circulation which is an indication of cellular leakage, loss of functional integrity of the cell membrane and necrosis in the liver (He and Aoyama, 2003) and the rise in the levels of serum total bilirubin (TB) is the most sensitive tool that reflects the severity of jaundice (Sturgill and Lambert, 1997). So, the degree and type of hepatocellular damage are evaluated by level of numerous above mentioned biochemical parameters in circulation, along with histological assessment of liver sections. Thus the alleviation in serum enzyme levels of a drug towards respective normal values, which were raised by a hepatotoxin, is an unambiguous sign of its hepatoprotective effects. In our study, CCl₄ treated group has highly raised levels of serum enzyme markers (SGOT, SGPT and ALP) along with damaged liver architecture. The SS extract (500 mg/kg) was found to be produced more significant hepatoprotection, both structurally and functionally as compared to both Silymarin (100 mg/kg) and SS extract (750 mg/kg). Decreased levels of SGOT and SGPT seem to protect the structural integrity of the hepatocellular membrane or accelerated regeneration/repairing of damaged hepatocytes produced by CCl₄, while decreased ALP and TB levels proposed the constancy of the biliary functions during damage with CCl₄. According to phytochemical analysis, SS Berries contain solanocarpine, solanocarpidine (Qureshi, 2004). Berries and leaves contain solasodine (Maiti et al., 1979). Plant also contains glycosides such as spirosolane, spirostane, furostane, diosgenin, pregnane (Nohara et al., 2007), and β-solanargine (Jawahar et al., 2004). In

addition, steroidal glycoside indioside D, triterpenoid glycosides, alkaloids, iridoids, flavonoids, lignans and quinines are also reported (Haribal et al., 2006). Vitamin C, α- and β-carotenes are present in *Solanum nigrum* (Sultana et al., 1995). So there is also a possibility that Vitamin C, α- and β-carotenes may also be present in other species of *Solanum*. The observed protective effects of SS ethanolic extract against CCl₄ induced liver damage might be due to the presence of these polyphenolic compounds (flavonoids, quercetin etc...), carotenoids, lignans, quinines, Vitamin C and steroid glycosides among other plant constituents. Phenolic compounds amongst many other constituents have been shown to possess hepatoprotective and calcium antagonist activities (Khalid et al., 2002) and the presence of such constituents in the extract, may be responsible for some of the pharmacological activities observed in this study. Another possibility is that SS may prevent lipid peroxidation by metal chelation and increased electron trapping capability probably due to the concurrent occurrence of both quinine skeleton and phenolic groups (Shiang et al., 1995). It is reported that the mice knocked out of *CYP2E1* gene show resistance against CCl₄ induced hepatotoxicity and the level of reactive metabolites can be reduced by inhibition of *CYP2E1* gene expression, consequently tissue injury is reduced (Wong et al., 1998). In recent years, there has been an active search for the development of *CYP₄₅₀* inhibitors from natural products that may have therapeutic potential in prevention of liver damage. Triterpene acids, oleanolic acid and ursolic acid inhibit *CYP₄₅₀* (Kim et al., 2004). So, the hepatoprotective action of SS extract may be due to the presence of some of the above mentioned compounds which cause down regulation of *CYP2E1* gene expression but it must be confirmed after a detail phytochemical analysis of the plant. To be brief, the possible hepatoprotective mechanism of SS aerial parts ethanolic extract (500 mg/kg) on CCl₄-induced liver injuries may be through one of the following actions;

- 1- Prevention of process of lipid per oxidation.
- 2- Free radical scavengers.
- 3- Down regulation of *CYP2E1* gene expression.

In conclusion, our present study offers scientific basis for the traditional use of *Solanum surattense* in hepatobiliary diseases in Eastern

system of medicine, particularly at a dose of 500 mg/kg. However, higher concentrations are less effective. It is suggested that further studies should be carried out to determine the therapeutic index and exact mechanism of hepatoprotection offered by the plant.

Table 1
Effects of ethanolic extract of SS (aerial parts) on rabbits serum biochemical parameters after CCl₄ administration

Group	SGOT (IU/l)	SGPT (IU/l)	ALP (IU/l)	TB (mg/dl)	Liver damage (Histological scores)
Normal control (5 ml/kg Normal saline)	40.69 ± 19.94	41.66 ± 23.35	264.5 ± 49.72	0.83 ± 0.22	0
CCl ₄ control (5 ml/kg Normal saline + 0.75 ml/kg)	455.2 ± 37.12*	434.2 ± 34.30*	394.3 ± 29.56*	1.32 ± 0.20	+3
Silymarin control (100 mg/kg + CCl ₄)	176.5 ± 56.77*°	205.9 ± 36.59*°	257.0 ± 41.03°	1.01 ± 0.42	+1
Test group 1 <i>Solanum surattense</i> extract (500 mg/kg + CCl ₄)	82.40 ± 26.85°	82.38 ± 20.10°	120.0 ± 38.06*°	1.28 ± 0.17	0
Test group 2 <i>Solanum surattense</i> extract (750 mg/kg + CCl ₄)	166.8 ± 27.66*°	333.2 ± 36.24*	348.3 ± 48.06	1.54 ± 0.04*	+2

Values are represented as Mean ± S.E.M. (n=10). 0 = Normal. +1 = Mild. +2 = Moderate. +3 = Severe.

* P < 0.05 compared with normal control group. ° P < 0.05 compared with CCl₄ control group.

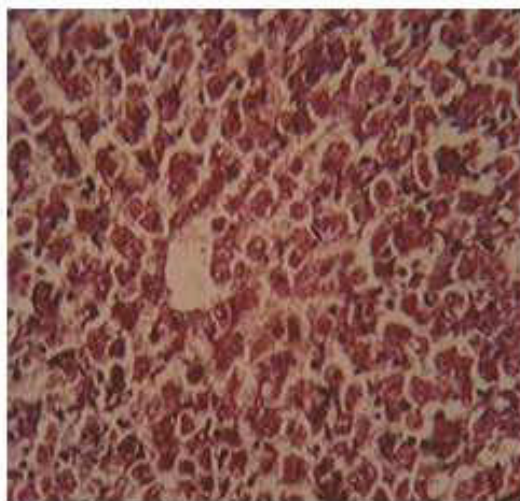


Figure A

A microphotograph of histopathological examination of randomly selected, formalin fixed, paraffin embedded, H & E-stained liver section of rabbit from Normal control group (Normal saline). Liver section shows normal liver morphology; Hepatocytes have round nucleus with centrally plus homogenous cytoplasm, flat endothelial cells around central vein and sinusoid.

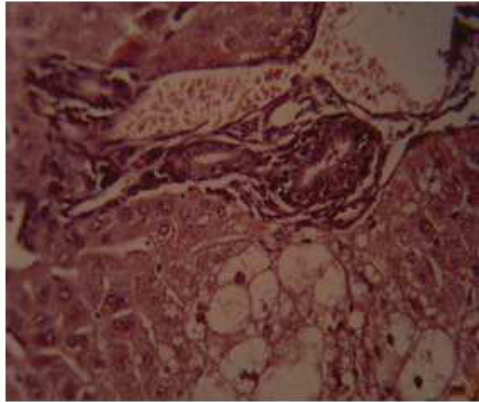


Figure B

A microphotograph of histopathological examination of randomly selected, formalin fixed, paraffin embedded, H &E-stained liver section of rabbit from CCl₄ control group (Normal saline + CCl₄). In liver section, 3-4 hepatocytes rows around central vein demonstrated; hepatocytes degeneration and necrosis, degeneration cells, endothelial lining of central vein showed more cell damage increased fat vacuoles in hepatocytes than level +2, focal necrosis and Bile duct proliferation.

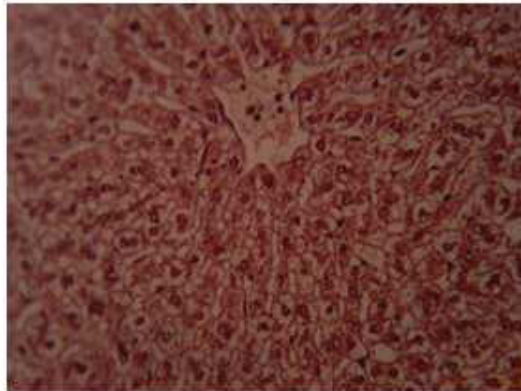


Figure C

A microphotograph of histopathological examination of randomly selected, formalin fixed, paraffin embedded, H &E-stained liver section of rabbit from Silymarin control group (100mg/kg + CCl₄). In liver section, 1-2 hepatocytes rows around central vein showed; hepatic cell degeneration along with necrosis (loss of nucleus), less injury of endothelial cells around central vein and less fat vacuoles in hepatocytes

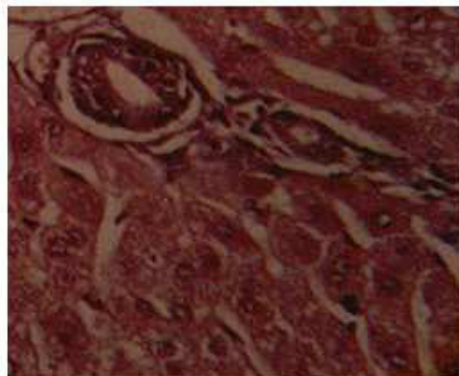


Figure D

A microphotograph of histopathological examination of randomly selected, formalin fixed, paraffin embedded, H &E-stained liver section of rabbit from Test group 1 (Solanum 500mg/kg + CCl₄). Liver section shows normal liver morphology; Hepatocytes have round nucleus with centrally plus homogenous cytoplasm, flat endothelial cells around central vein and sinusoid.

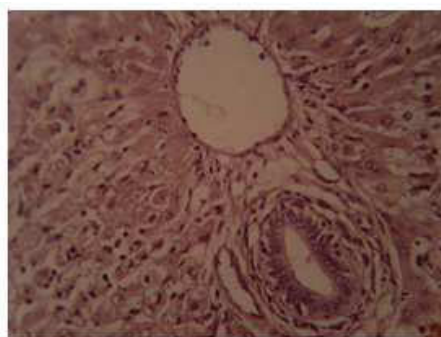


Figure E

A microphotograph of histopathological examination of randomly selected, formalin fixed, paraffin embedded, H &E-stained liver section of rabbit from Test group 2 (*Solanum* 750mg/kg + CCl_4). In liver section, some hepatocytes rows around central vein showed; swelling, intracytoplasmic vacuolar degeneration in centrilobular, midzonal and periportal areas, endothelial cells around central vein more damage than level +1, more fat vacuoles in hepatocytes than level +1.

ACKNOWLEDGEMENTS

The authors are indebted to Dr. Mohammad Arshad from Cholistan Institute of Desert Studies, the Islamia University of Bahawalpur, Pakistan, for providing expertise in identification of plants and also to Dr. Khalid Usman, Associate Professor, Pathology Department, Quaid-e-Azam Medical College, Bahawalpur, Pakistan, for providing his expertise in carrying out histopathological studies.

REFERENCES

1. Baig MS, Akram M, Hassan MA. Possibilities for range development in Cholistan desert as reflected by its physiography and soils. *Pakistan Journal of Biological Sciences* 1980;30:61-71.
2. Chouhan F, Wattoo MHS, Tirmzi SA, Memon FZ, Aziz-ur-Rehman, Tufail M. Analytical investigation of inorganic nutritive elements of *Capparis decidua* grown in Cholistan desert [Pakistan]. *The Nucleus* 2002;39(3-4):195-199.
3. Dabur R, Gupta A, Mandal TK, Singh DD, Bajpai V, Gurav AM, Lavekar GS. Antimicrobial activity of some Indian Medicinal Plants. *African Journal of Traditional, Complementary and Alternative Medicines* 2007;4(3):313-318.
4. Haribal M, Renwick JAA, Attygalle AB, Kiemle D. A Feeding Stimulant for *Manduca sexta* from *Solanum surattenses*. *Journal of Chemical Ecology* 2006;32: 2687–2694.
5. He G, Aoyama Y. Effects of adding some dietary fibers to a cystine diet on the activities of liver antioxidant enzymes and serum enzymes in rats. *Bioscience, Biotechnology, Biochemistry* 2003;67:617-621.
6. Humason GL. 1979. Animal Tissue Techniques. W. H. Freeman and Company, San Francisco.
7. Jawahar M, Rabert GA, Jeyaseelan M. Rapid Proliferation of Multiple Shoots in *Solanum trilobatum* L. *Plant Tissue Culture* 2004;14(2):107-112.
8. Khalid HJ, Sheikh AS, Gilani AH. Protective effect of rutin on paracetamol and CCl_4 induced hepatotoxicity in rodents. *Fitoterapia* 2002;73:557-563.
9. Kim KA, Lee JS, Park HJ, Kim JW, Kim CJ, Shim IS, Kim NJ, Han SM, Lim S. Inhibition of cytochrome P450 activities by oleanolic acid and ursolic in human liver microsomes. *Life Sciences* 2004;74:2769-2779.

10. Maiti PC, Mookherjea S, Mathew R, Dan SS. Studies on Indian *Solanum l.* alkaloid content and detection of Solasodine. *Economic Botany* 1979;33(1):75-77.
11. Nohara T, Ikeda T, Fujiwara Y, Matsushita S, Noguchi E, Yoshimitsu H, Ono M. Physiological functions of Solanaceous and tomato steroidal glycosides. *Journal of Natural Medicine* 2007;61:1-13.
12. Qureshi R. 2004. Floristic and ethnobotanical study of Desert-Nara Region, Sindh [dissertation]. Shah Abdul-latif University, Khairpur, Sindh, Pakistan.
13. Shafi MS, Ashraf MY, Sarwar G. Wild medicinal plants of Cholistan area of Pakistan. *Pakistan Journal of Biological Sciences* 2001;4(1):112-116.
14. Shiang SH, Sheau FY, Chaunge YH. Effect of anthraquinone derivatives on lipid peroxidation in rat heart mitochondria: structure-activity relationship. *Journal of Natural Products* 1995;58:1365-1371.
15. Sturgill MG, Lambert GH, 1997. Xenobiotic-induced hepatotoxicity; mechanisms of liver injury and methods of monitoring hepatic function. *Clinical chemistry* 43, 1512-1526.
16. Sultana, S., Shahid, P., Mohammad, I., Mohammad, A., 1995. Crude extracts of hepatoprotective plants, *Solanum nigrum* and *Chichorium intybus* inhibits free radical mediated DNA damage. *Journal of Ethnopharmacology* 45, 189-192.
17. Weber, L., Boll, W.D., Stampfl, A., 2003. Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model. *Critical Reviews in Toxicology* 33, 105-136.
18. Wong, F.W., Chan, W.Y., Lee, S.S., 1998. Resistance to carbon tetrachloride-induced hepatotoxicity in mice which lack *CYP2E1* expression. *Toxicology and Applied Pharmacology* 153, 109-118.